

Soil health and pesticide effects

Kees van Gestel

Department of Ecological Science,
Vrije Universiteit,
Amsterdam



The need for sustainable soil use (Wall & Six, 2015)



U.S. President Franklin D. Roosevelt (1937):
“A nation that destroys its soils destroys itself.”

“We can’t breathe, eat, drink, or be healthy without sustainably managing soils.”



Effects of land-use intensification (Tsiafouli et al. 2015)

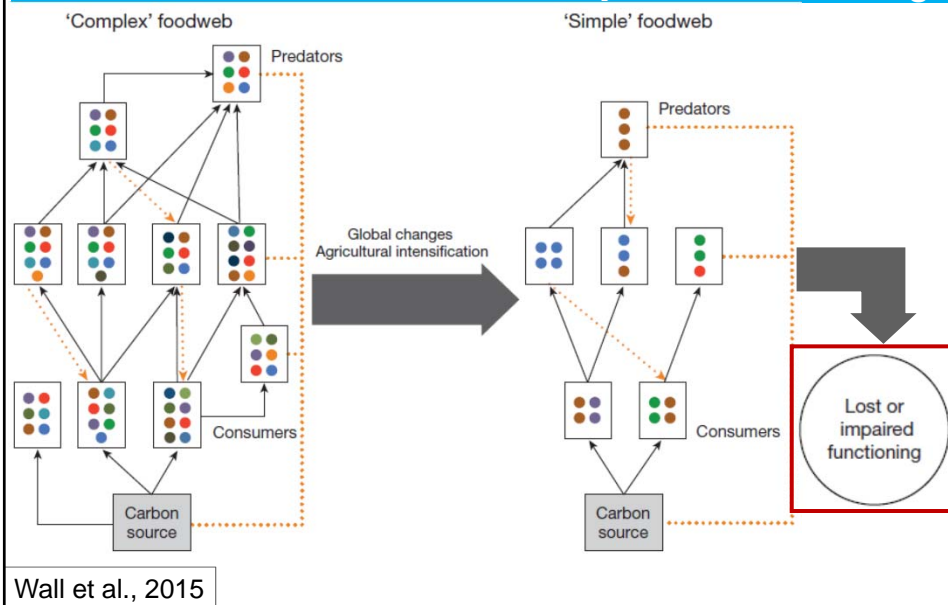
Increasing land-use intensity **reduced**

- ▲ complexity of **soil food webs**
- ▲ community-weighted mean **body mass** of soil fauna
- ▲ **species richness** of earthworms, Collembolans, oribatid mites
- ▲ etc.

Overall effects of land-use intensification:


- ▲ soil food webs **less diverse**, composed of **smaller organisms**.
- ▲ **fewer functional groups** of soil biota with **fewer and taxonomically more closely related species**

Key features in reduction of species in soil food webs - adverse effect on ecosystem functioning

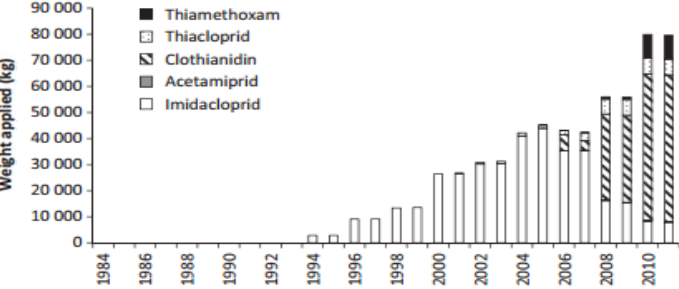


Neonicotinoids

- Introduced as insecticides in 1990 (Kollmeyer *et al.* 1999)
- Agricultural use → seed dressing and foliar sprays (Goulson 2013)
- Systemic → **whole plant is protected** (Goulson 2013)
- Sunflower, maize, rape seed, cereals and potatoes (Goulson 2013)
- Application rates 30-150 g/ha (~0.03-0.15 mg/kg in top soil)




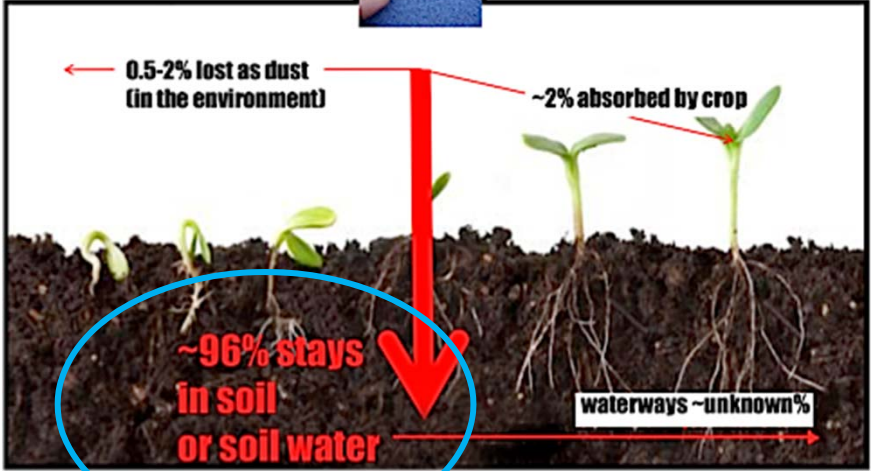
**60%
seed
dressing
treatment**



Environmental fate of neonic seed dressings

(Source: Goulson)

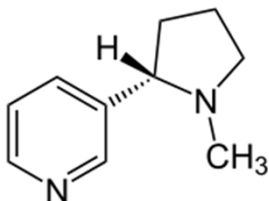




Goulson 2013

Origin/chemical properties

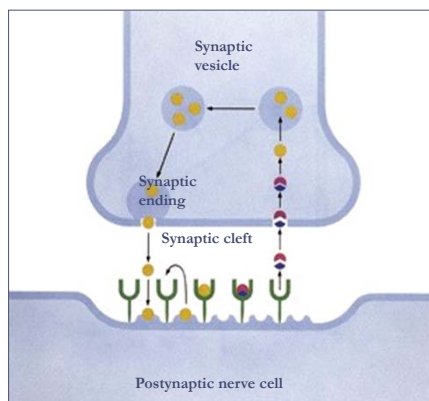
- Based on known action of nicotine as natural insecticide
- Nicotine is alkaloid acting on acetylcholine receptor



- 7 Neonicotinoids on the market;
- Two main chemical groups (nitro \leftrightarrow cyano substituted)

Mode of Action

- Bind irreversibly to nicotinic acetylcholine receptor on post-synaptic membrane
- Act as acetylcholine agonist \rightarrow Paralysis or death



High receptor binding affinity:

- \rightarrow Low-dose exposure over extended periods of time can culminate into substantial effects
- \rightarrow Limits potential for recovery

Recent reviews on Neonicotinoids

(Introduced by Van der Sluijs et al., 2015)

- **Persistent** (for several compounds $T_{1/2} \gg 100$ days)
 - Reasonably **soluble** in water → potential **transport** in soil
 - Some **metabolites** also highly toxic
- May accumulate upon repeated application
- Widely distributed in environment
- Concentrations in soil typically in $\mu\text{g}/\text{kg}$ – mg/kg range
- Often mixtures of active substances and metabolites

Recent reviews on Neonicotinoids

(Van der Sluijs et al., 2015; Pisa et al., 2015; Chagnon et al., 2015)

Knowledge gaps include e.g.:

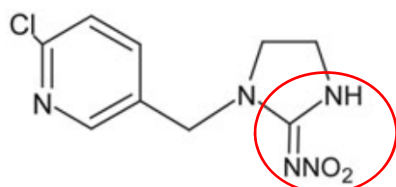
- Fate in soil
- Toxicity to soil organisms beyond earthworms
- Long-term effects
- Mixture toxicity
 - neonicotinoids and metabolites
 - Interaction with other types of pesticides
- Effects on ecosystem services in soil (due to effects on earthworms and springtails)

Two compounds; two chemical classes

Imidacloprid

- First (1991)
- >120 Countries; >140 crops.
- Highly toxic to honey bees and also for aquatic insects.
- **EU ban from December 2013**

N-nitroguanidines → nitro-substituted

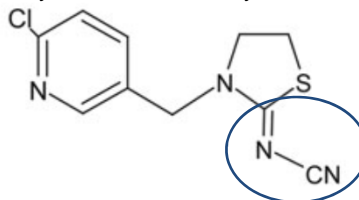


LD₅₀ 0.018 µg/bee

Thiacloprid

- New type (2000)
- Agricultural crops, esp. pome fruits
- Same molecular mechanism but significantly different responses

N-cyano-amidines → cyano substituted.



LD₅₀ 14.6 µg/bee

Iwasa et al. 2004

Research Questions - 1



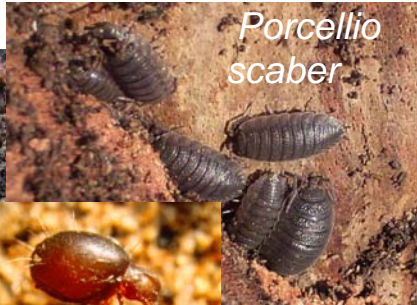
Toxicity of neonicotinoids to different soil invertebrate species

1. How toxic are imidacloprid and thiacloprid to soil invertebrates?
2. Which soil invertebrate species are most sensitive to imidacloprid and thiacloprid?
2. Do imidacloprid and thiacloprid have similar toxicity to soil invertebrates?

Test organisms



Eisenia andrei



Porcellio scaber



Oppia nitens



Enchytraeus crypticus



Folsomia candida

Methods - general

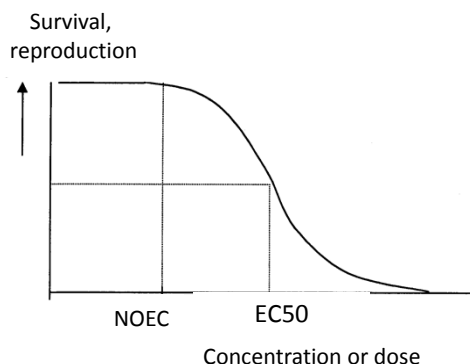
- LUFA 2.2 natural soil
- Treatments: range of insecticide concentrations in soil
- Controls
- Climate chamber; 20 °C, 16 hrs dark: 8 hrs light; 75% RH
- 21-28 days exposure
- Survival, reproduction (# juveniles), growth








Data analyses – dose-response

- Dose response models → LC_{50} , EC_{50} , etc.

(Hamilton et al. 1977; Haanstra et al. 1985)



Summary of results

Species	Imidacloprid values mg/kg dry soil		Thiacloprid values mg/kg dry soil	
	LC_{50}	EC_{50}	LC_{50}	EC_{50}
<i>F. candida</i> 	0.20 – 0.64	0.10 – 0.26	2.7 – 3.9	1.7 – 2.4
<i>E. andrei</i> 	0.77	0.39	7.1	0.44
<i>E. crypticus</i> 	>30	2.0	>30	12
<i>P. scaber</i> 	7.6	6.7	>32	>32
<i>O. nitens</i> 	360	119	>1000	>100

De Lima e Silva et al. 2017

Research Questions - 2

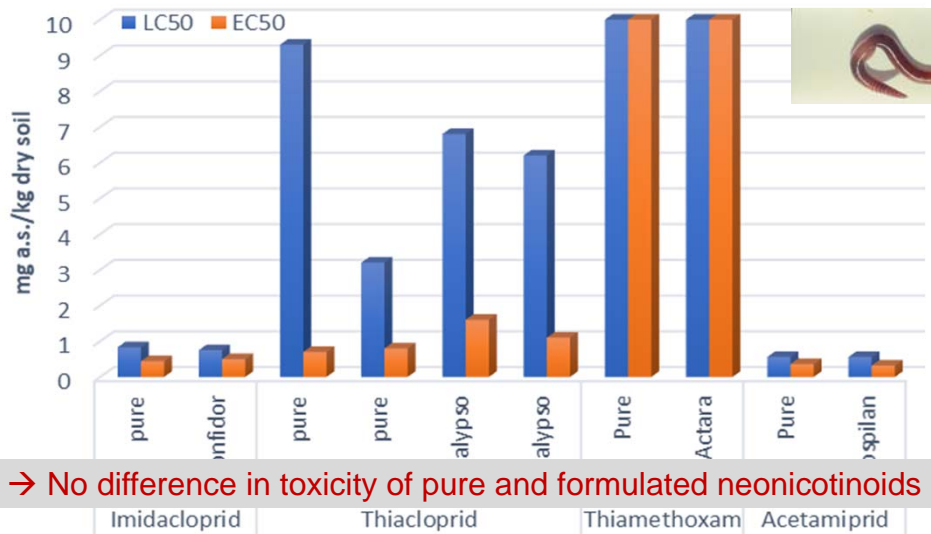
Starting points:

- All pesticides applied in **commercial formulations**
- Formulations contain several **additives** that might affect their efficacy, so also their toxicity

Question:

Are commercial formulations more (or less) toxic than pure active substances?

Toxicity of active substances ↔ formulations to earthworms *Eisenia andrei*



→ No difference in toxicity of pure and formulated neonicotinoids

De Lima e Silva et al. In prep

Research Questions - 3 Multi-generation toxicity testing


Starting points:

1. Springtails quite sensitive to neonicotinoids
2. Neonicotinoids bind irreversibly to acetyl choline receptor
→ **cumulation of effects expected** upon long-term exposure
3. Imidacloprid much more persistent than thiacloprid

Aims:

- Determining multi-generation effects of **imidacloprid** and **thiacloprid** to *Folsomia candida* in soil spiked once
- Multigeneration toxicity of **thiamethoxam** to *Folsomia candida* in repeatedly spiked soil

Multi-generation tests *Folsomia candida*



Imidacloprid

Generation	EC50 (mg/kg dry soil)
P	0,29
F1	0,12
F2	0,14

Imidacloprid; $T_{1/2} > 125$ days

Thiamethoxam

Generation	Pure	Actara*
P	0,23	0,25
F1	0,23	0,16
F2	0,3	>0,37

Thiamethoxam; $T_{1/2} = 140-165$ days (pure)
 $T_{1/2} = 46-74$ days (Actara*)

Thiacloprid

Generation	EC50 (mg/kg dry soil)
P	1,5
F1	>3,3
F2	>3,3

Thiacloprid; $T_{1/2} = 10-12$ days

Imidacloprid quite stable: $T_{1/2} > 125$ days
Thiamethoxam repeated dosing + quite stable: $T_{1/2}$ 46-165 days
 → No toxicity change upon long-term incubation
Thiacloprid: decrease in toxicity after first generation
 → agrees with fast degradation ($T_{1/2}$ ~10-12 days)

Van Gestel et al. 2017; De Lima e Silva et al. In press

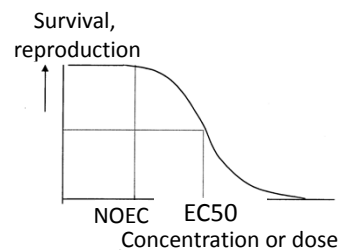
Answers to research questions

1. Toxicity of imidacloprid and thiacloprid to soil invertebrates?
 - Springtails (*Folsomia candida*) most sensitive
 - Earthworms (*Eisenia andrei*) also quite sensitive
2. Differences in the toxicity of imidacloprid and thiacloprid?
 - For survival in some cases considerable differences
 - Little difference for sub-lethal effects
 - Suggests difference in mode of action
3. Toxicity of formulated products versus active substances?
 - No increased toxicity of compounds in formulation
4. Long-term toxicity?

Toxicity of imidacloprid & thiamethoxam to springtails did not change upon long-term incubation; thiacloprid toxicity decreased due to fast degradation

Risk of neonicotinoids to soil invertebrates

General paradigm:
dose determines effect



Risk assessment

→ compare exposure and effect concentrations

Risk assessment of neonicotinoids to soil invertebrates

- **Exposure:** Predicted soil concentration (top 5 cm layer):
0.03-0.15 mg/kg
 - **Effect:** EC₅₀s 0.10-0.44 mg/kg (springtails, earthworms)
- Soil concentrations close to or above effect concentrations, already after single application
- Potential risk to soil invertebrates of neonicotinoids studied

Conclusion – remaining questions

Neonicotinoids not only harmful for honey bees but also for soil invertebrates

- Several questions remain regarding
 - Exposure under field conditions?
 - Why difference in toxicity of imidacloprid and thiacloprid?
 - Role of toxicokinetics?
 - Difference in response of earthworms and enchytraeids?
 - How to explain high sensitivity of earthworms?
 - Possible ecosystem effects (biodiversity)?
 - Potential risk for soil ecosystem services? } Soil health
 - Etc.

Acknowledgements

- Claudia de Lima e Silva (PhD student)
- Nicola Brennan, Justine Marriette, Jan Kok (MSc students),
Thao Lam, Daniël Commandeur, Jitske Brouwer (BSc students)
- Rudo Verweij for continuous support of students with all aspects
of the experimental work
- Jacco Koekkoek and Marja Lamoree (VU Dept. Environment &
Health) for chemical analysis of neonicotinoids in soil
- Bayer Crop Sciences for providing test chemicals

